

Automated Bladder Cancer Screening with Deep Learning Algorithms

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ABSTRACT

- Background: Bladder cancer has a high recurrence rate, requiring lifelong surveillance. Urine cytology is noninvasive but labor-intensive and subject to variability in assessing nuclear-to-cytoplasmic (N/C) ratio.
- Objective: Develop an automated segmentation pipeline to objectively measure N/C ratio in urine cytology images.
- Methods: Trained a U-Net with a ResNet-50 encoder on 300 urothelial cell images with ground-truth nucleus and cytoplasm masks and compared performance with Otsu thresholding.
- Results: U-Net achieved a Spearman correlation coefficient of 0.844 with the ground-truth N/C ratios, compared with 0.765 for Otsu thresholding. Otsu failed to detect nuclei in 9/200 cells.
- Conclusion: Mean N/C ratio increased across diagnostic categories (0.185 → 0.250 → 0.397 → 0.488), supporting N/C ratio as a quantitative biomarker and demonstrating the potential of deep learning for more reproducible bladder cancer screening.

INTRODUCTION

Challenges in Urine Cytology:

- Manual screening of urine cytology specimens is time consuming and laborious, can also lead to subjective N/C ratio estimates, hurting reproducibility
- Overlapping cells, debris, and low contrast break simple thresholding rules, can lead to misclassification

Intended Solutions & Next Steps:

- Train a U-Net model to segment the nucleus, cytoplasm, and background of cell image while also computing per cell N/C ratio
- Use a simple baseline such as Otsu thresholding for comparison and interpretability of per cell N/C ratio across the four diagnostic categories (atypical suspicious, positive, negative)
- Combine N/C ratios across cells for the specimen for diagnosis support

Data & Evaluation Plan:

- Urothelial cell dataset: 200 with ground truth/cytoplasm masks
- Specimen dataset: 25 cells/patients across diagnostic categories
- Grayscale preprocessing
- Compared U-NET and Otsu Method
- Evaluated using Dice score and Spearman coefficient correlation
- Analyzed N/C ratio across diagnostic categories

METHODS

Datasets

Urothelial Cell Dataset (model training):

- 300 urothelial cells, each with ground truth segmentation masks for nucleus and cytoplasm
- Used to train and benchmark segmentation approaches, and to calculate the nucleus-to-cytoplasm (N/C) ratio

Specimen Cell Dataset:

- Contains 25 cells per patient, grouped by diagnostic category
- Used to investigate whether the calculated N/C ration can be used as a predictive biomarker across patient diagnosis

Preprocessing, Models, Evaluation Metrics

Preprocessing:

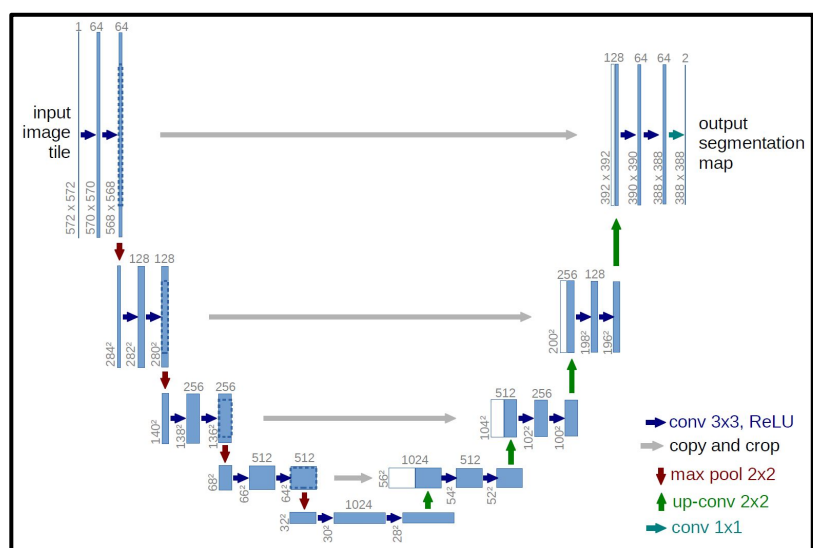
- Grayscale conversion: Converted urothelial dataset images to grayscale for better intensity based analysis and to establish a baseline for N/C ratio prediction

Models:

- Otsu's Method: Segmentation Approach
- UNET: Deep Learning Approach

Evaluation Metrics:

- Dice Score Coefficient
- Spearman correlation
- Analyzed N/C ratio variation by diagnosis



UNET

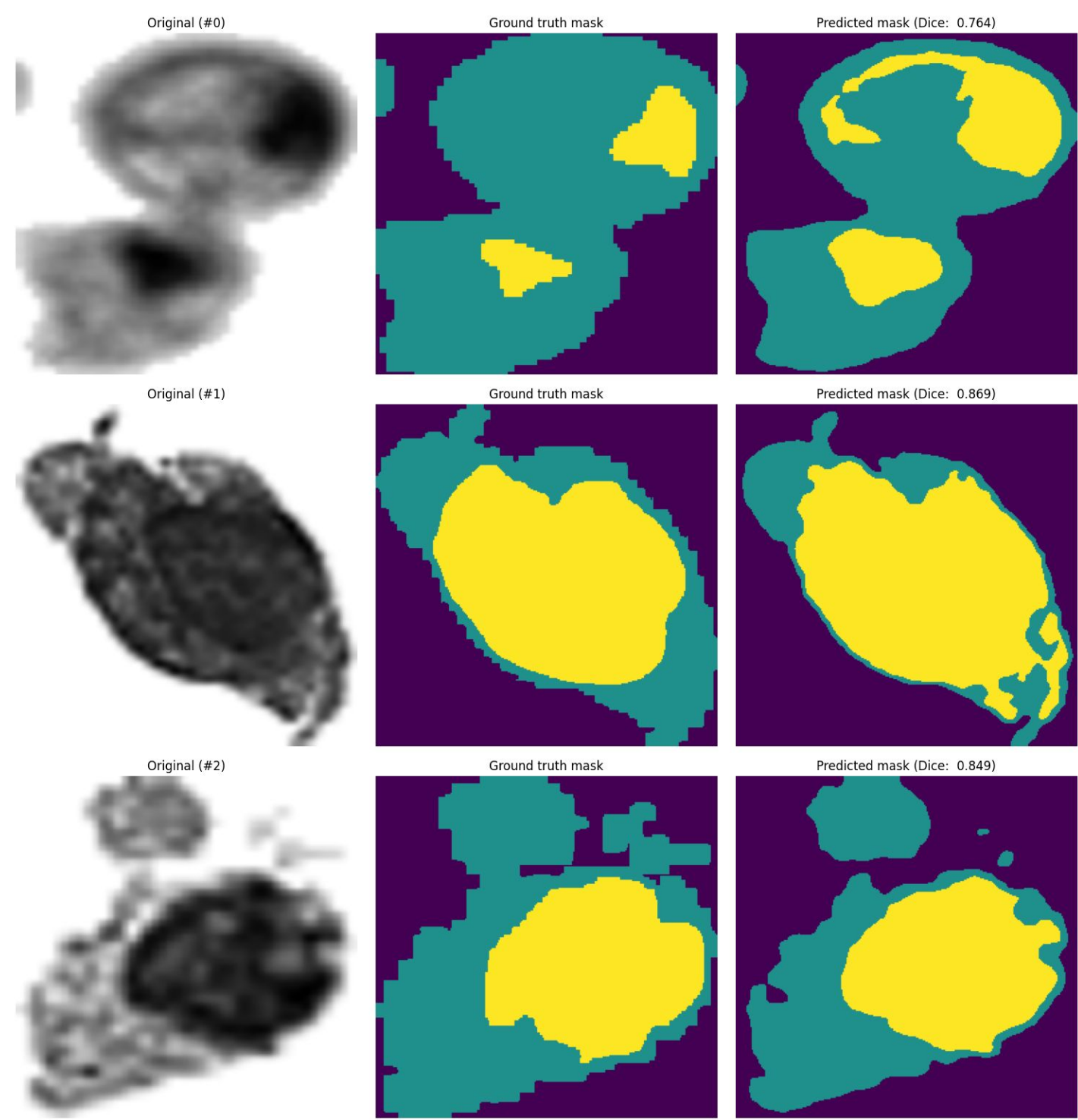
Otsu's Method:

- Classes: 3 (multi-Otsu thresholding: background, cytoplasm, nucleus)
- Preprocessing: grayscale conversion, intensity rescaling to 0-1, Gaussian smoothing ($\sigma = 1.5$, tuned via grid search), inverted so nuclei have higher intensity
- Post-processing: fill small holes in segmented regions (area threshold = 200 pixels) to clean up noisy boundaries
- N/C ratio = nucleus pixels / (nucleus + cytoplasm pixels)

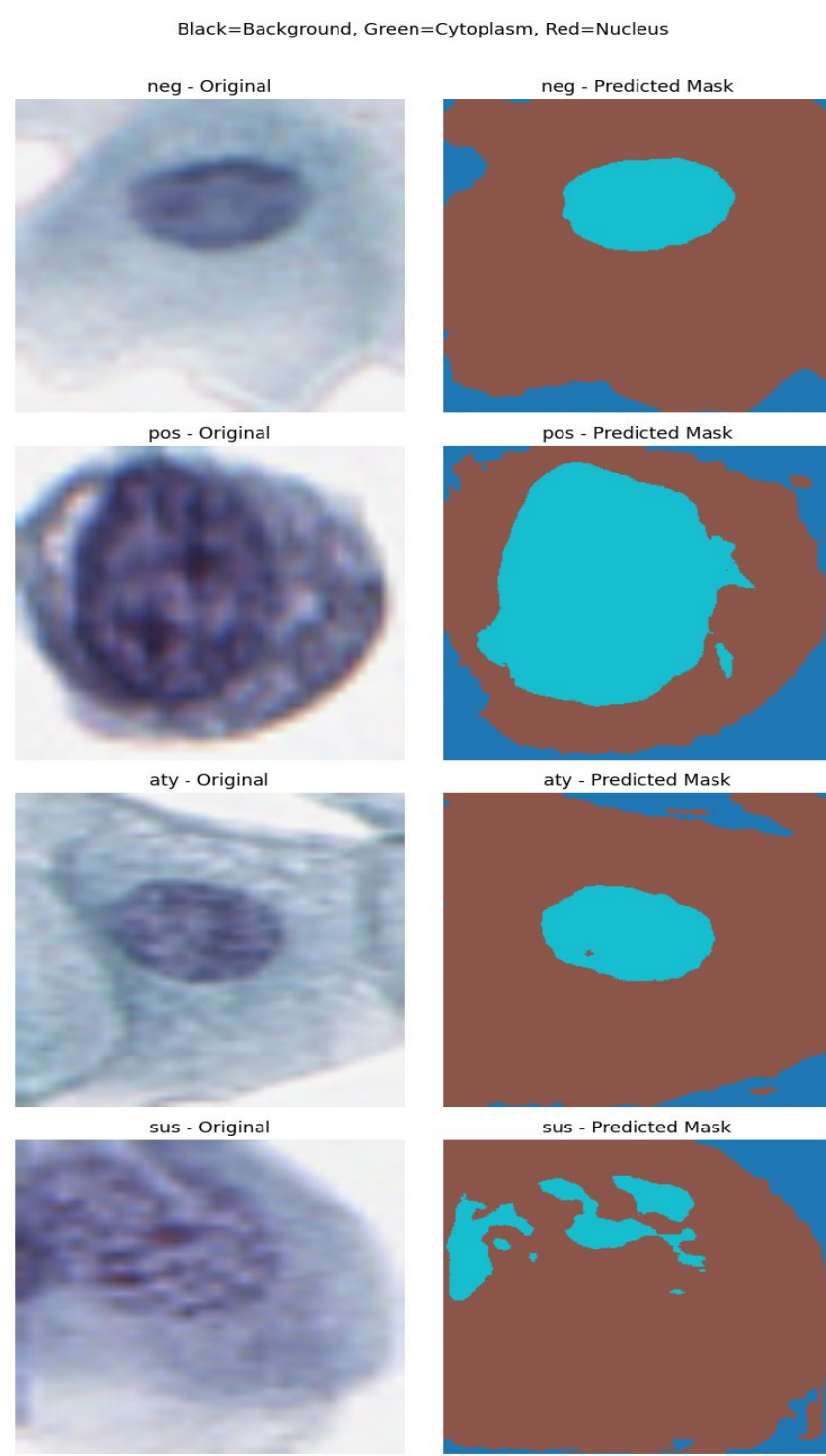
UNET:

- Epochs: 50
- Batch size: 8
- Loss function: 50% Cross_Entropy_Loss + 50% Dice Loss
- Optimizer: Adam, learning rate 1e-4
- N/C ratio = nucleus pixels / (nucleus + cytoplasm pixels)
- Resize to 256×256, normalize 0–1, channels-first conversion
- 3 classes for segmentation: background, cytoplasm, nucleus

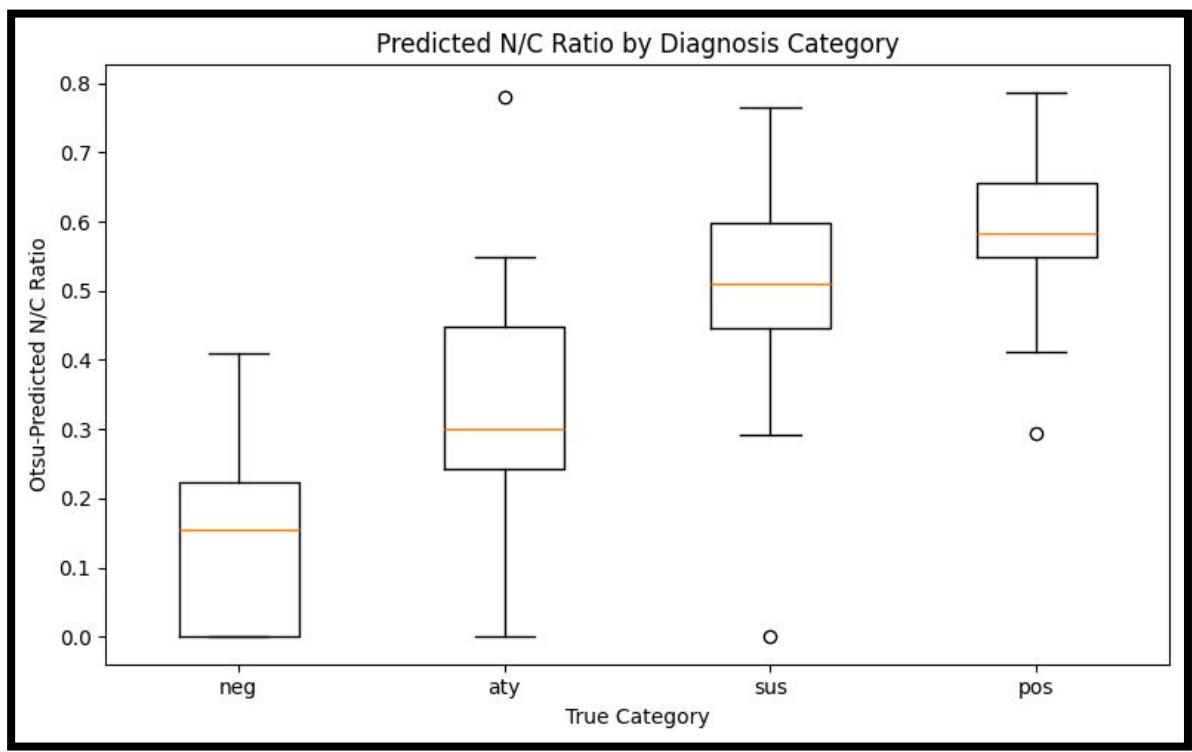
RESULTS



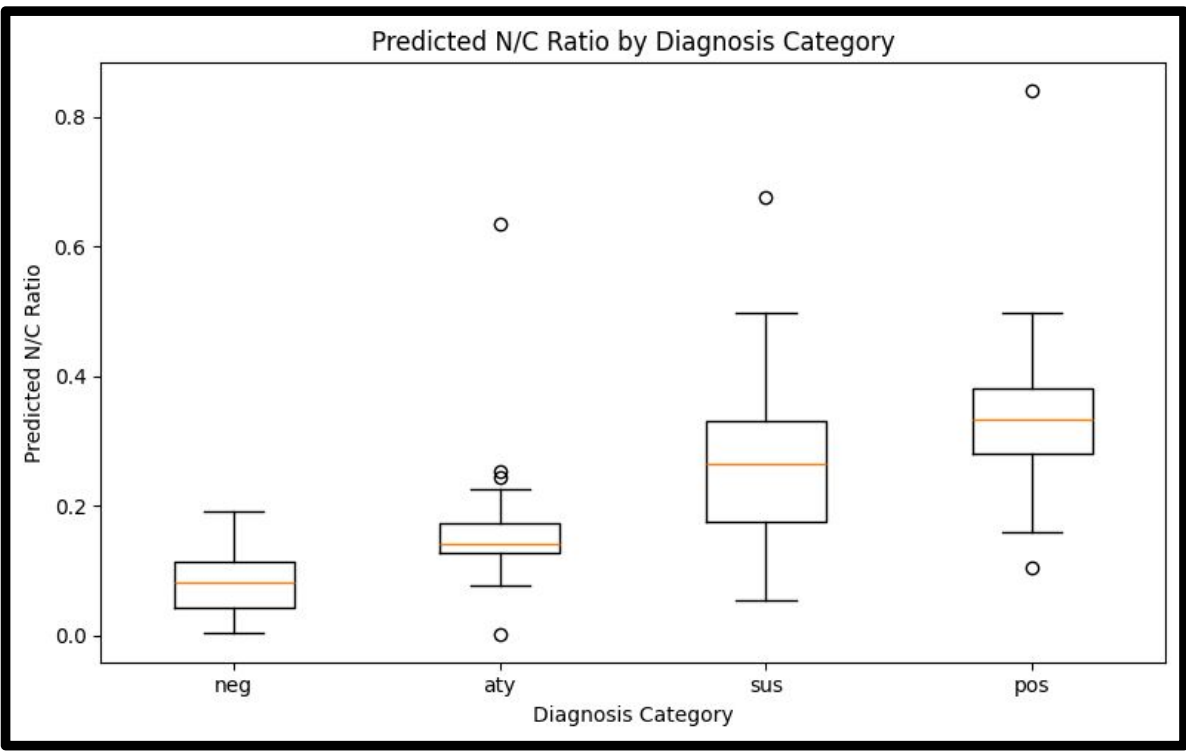
Otsu's Method mask
Original image, ground truth, and Otsu-predicted masks



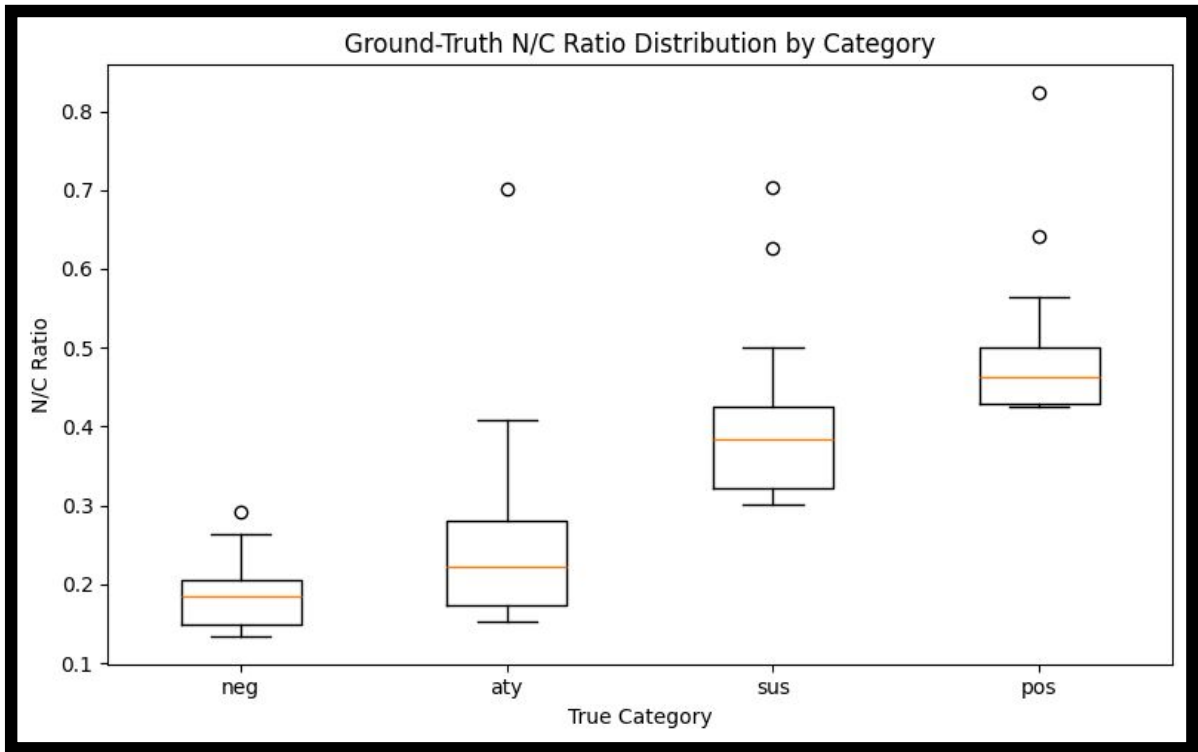
UNET mask
Original image and UNET-predicted masks



Otsu's Method: SCC = 0.765



UNET: SCC = 0.844



Ground truth

CONCLUSION

Future Impact:

- Scale & Improve:** Expand to larger, multi-institution datasets and combine N/C ratio with additional cellular features for more robust predictions.
- Target Hard Cases:** Focus on atypical/suspicious cells and improve performance where pathologists have the most difficulty reaching agreement.
- Whole-Specimen Analysis:** Automate cell detection across entire specimens and generate patient-level screening scores.

Limitations:

- limited dataset size (~300 cells for training, 25 cells/patient validation), which limits generalizability, and causes there to be a risk of overfitting
- N/C ratio may be insufficient, bladder cancer diagnosis depends on other morphological features
- Image quality challenges could lead to segmentation errors, which could then lead to potential misclassification

Future Directions:

- Expand the Dataset:** Include more patients, cells, and diverse clinical data from multiple institutions.
- Improve Detection:** Refine segmentation and cell detection for crowded or atypical cells.
- Enhance Biomarkers:** Combine N/C ratio with nuclear texture, chromatin density, and shape features.
- Clinical Validation:** Test the model on independent datasets and evaluate its potential for real-world screening.

